A live attenuated Salmonella Typhimurium oral T cell vaccine against PD-L1 protects 100% of animals from a leukemia challenge



Sébastien Wieckowski¹, Heiko Smetak², Marco Springer³, Iris Kobl³, Amine A. Berkane⁴, Ming Wei⁴, Albrecht Meichle³, Klaus M. Breiner¹, Philipp Beckhove², Marc Mansour¹, Matthias Schroff¹, Heinz Lubenau³



¹VAXIMM AG, Basel, Switzerland; ²Regensburg Center for Interventional Immunology (RCI), Regensburg, Germany; ³VAXIMM GmbH, Mannheim, Germany; ⁴CellVax S.A.S., Romainville, France.

Background

VAXIMM's oral T-cell vaccine platform is based on the approved, live attenuated *Salmonella* Typhi strain Ty21a vaccine, which has been administered in millions of individuals for prophylactic vaccination against typhoid fever. This strain has been thoroughly studied, is safe and well tolerated. The bacteria are modified to deliver an eukaryotic expression plasmid, which encodes the genetic information of a specific target antigen¹.

VXM01 lead vaccine encodes vascular endothelium growth factor receptor 2 (VEGFR2) in order to evoke an immune response specifically directed against the tumor vasculature. It is currently in clinical development as a treatment for various solid cancer types. The murine analogue of VXM01 has shown consistent anti-angiogenic activity in different tumor models in several animal studies². An increase in tumor immune cell infiltration was recently shown. A proposed mechanism of action of VXM01 is described in **Figure 1**.

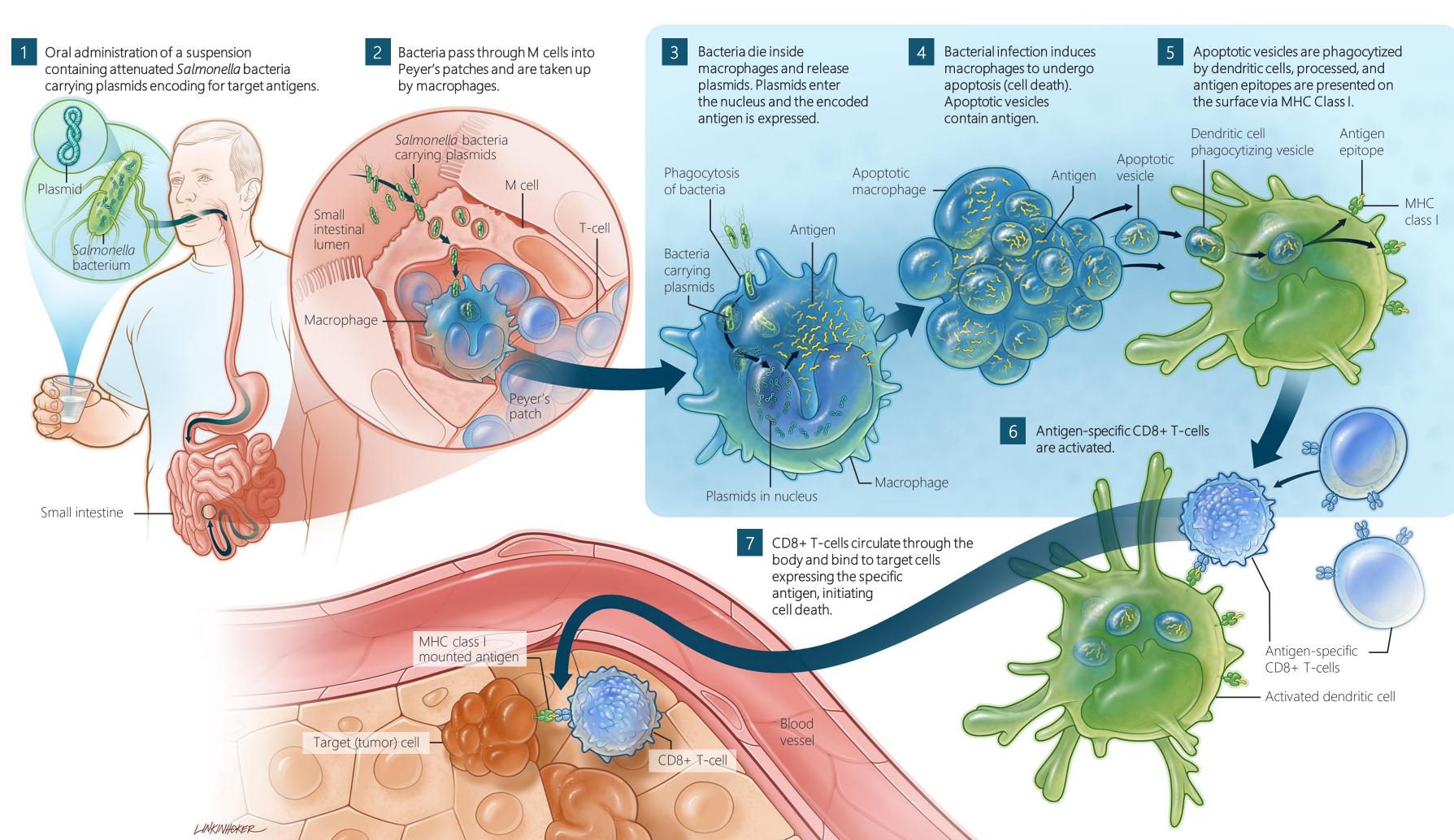


Figure 1. Intra-lymphatic delivery of *Salmonella* Typhi strain Ty21a T-cell vaccines via the oral route leading to target-specific T-cell activation.

current study summarizes the immunogenicity and preclinical anti-cancer efficacy for Salmonella Typhimurium SL7207 murine vaccines VXM10 (Figure VXM10a 2A), eukaryotic with transformed expression plasmids encoding the full-length murine programmed death-ligand 1 (PD-L1) protein and a truncated form of PD-L1, (Figure respectively 2B). Indeed, the deletion of the signal peptide (SP) prevents the proper localization of the native PD-L1 protein to the cell surface. The vector, i.e. without empty plasmid, was used as negative control throughout the study.

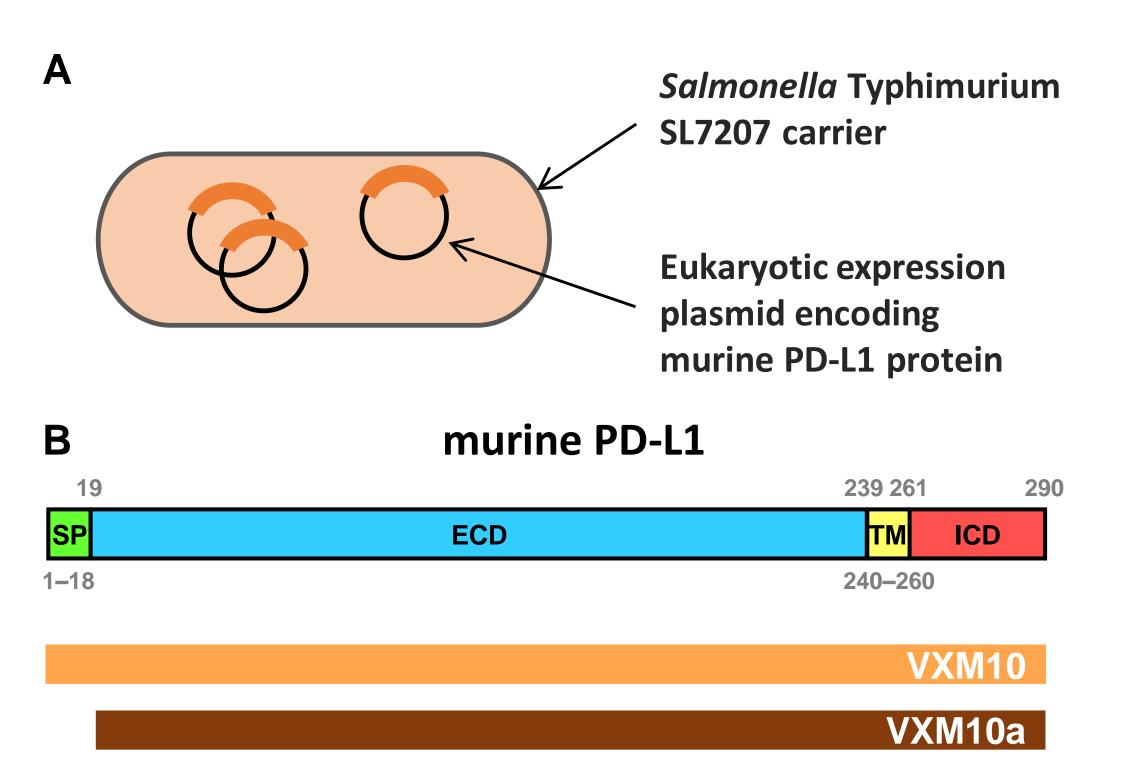


Figure 2. (A) Schematic representation of VXM10 oral T-cell vaccine, and (B) domains of the murine PD-L1 protein encoded in VXM10 (orange) and VXM10a (brown) vaccines.

Antitumor efficacy

We evaluated the prophylactic anti-cancer activity of VXM10 and VXM10a in the FBL-3 disseminated model of leukemia expressing PD-L1³ (**Figure 3A**). Empty vector, VXM10 and VXM10a were given by oral gavage at ca. 10^8 CFU and 10^{10} CFU, on days 1, 3, 5 and 7 as a prime vaccination, and on days 14 and 22 as boosts (**Figure 3B**). C57BL/6 mice (n=6 per group) then received 5×10^6 viable FBL-3 cells by intraperitoneal injection on day 20. All surviving animals were rechallenged with 5×10^6 viable FBL-3 cells by intraperitoneal injection on day 100.

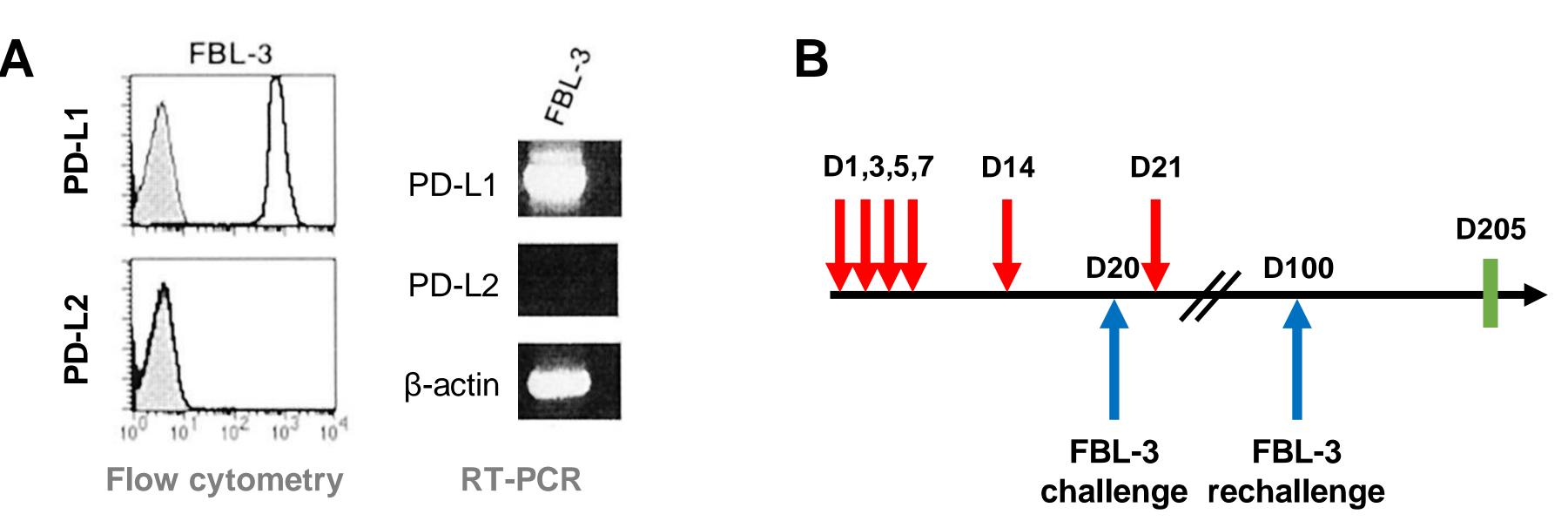


Figure 3. (A) Expression of PD-L1, but not PD-L2, by FBL-3 cell line³, as measured by flow cytometry (left inset) and RT-PCR (right), and (B) experimental design and treatment schedule in the prophylactic and re-challenge experiment.

Prophylactic vaccination with VXM10 and VXM10a was highly tolerated, as no deterioration in general status nor significant body weight loss were observed during the treatment (**Figure 4A**). It also generated a rapid and sustained anti-leukemia effect with 100% (6 out of 6) of surviving animals 80 days after leukemia challenge (P=0.0005) in the highest dose groups. In contrast, vaccination with the empty vector control did not show any anti-cancer activity, as the median survival reached 41 days, and 0% (0 out of 6) of cancer regression was observed (**Figure 4B**). Importantly, 100% of surviving mice in the high dose groups resisted rechallenge with FBL-3 cells for at least 100 days (P=0.0002), demonstrating that vaccination with VXM10 and VXM10a generated a potent memory T cell response against the leukemia (**Figure 4B**).

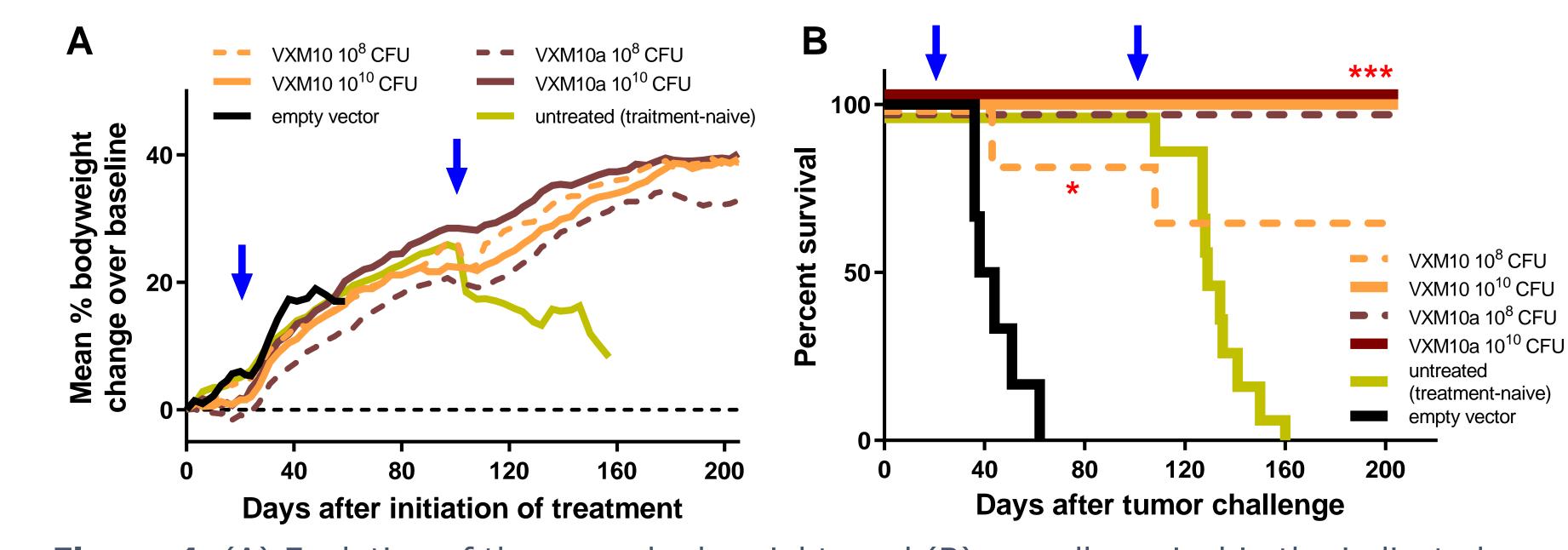


Figure 4. (A) Evolution of the mean bodyweight, and (B) overall survival in the indicated treatment groups and doses. The blue arrows represent the time points of leukemia challenge. Treatment-naive animals (yellow curves) were used as a control for the FBL-3 rechallenge and received the leukemia cells only day 100.

We finally evaluated the therapeutic efficacy of VXM10 and VXM10a in the FBL-3 model. C57BL/6 mice (n=8 per group) received 5×10^6 viable FBL-3 cells by intraperitoneal injection on day 0. Empty vector, VXM10 and VXM10a were then administered by oral gavage at a dose of 10^9 CFU on days 1, 3, 5 and 7 as a prime vaccination, and on days 14 and 21 as boosts (**Figure 5**).



Figure 5. Experimental design of the therapeutic study.

Therapeutic vaccination with VXM10 and VXM10a was well tolerated (**Figure 6A**), and induced full leukemia control, with 100% (8 out of 8) of surviving animals 94 days after leukemia challenge (P<0.0001). In contrast, treatment with the empty vector control did not show any anti-cancer effect (**Figure 6B**).

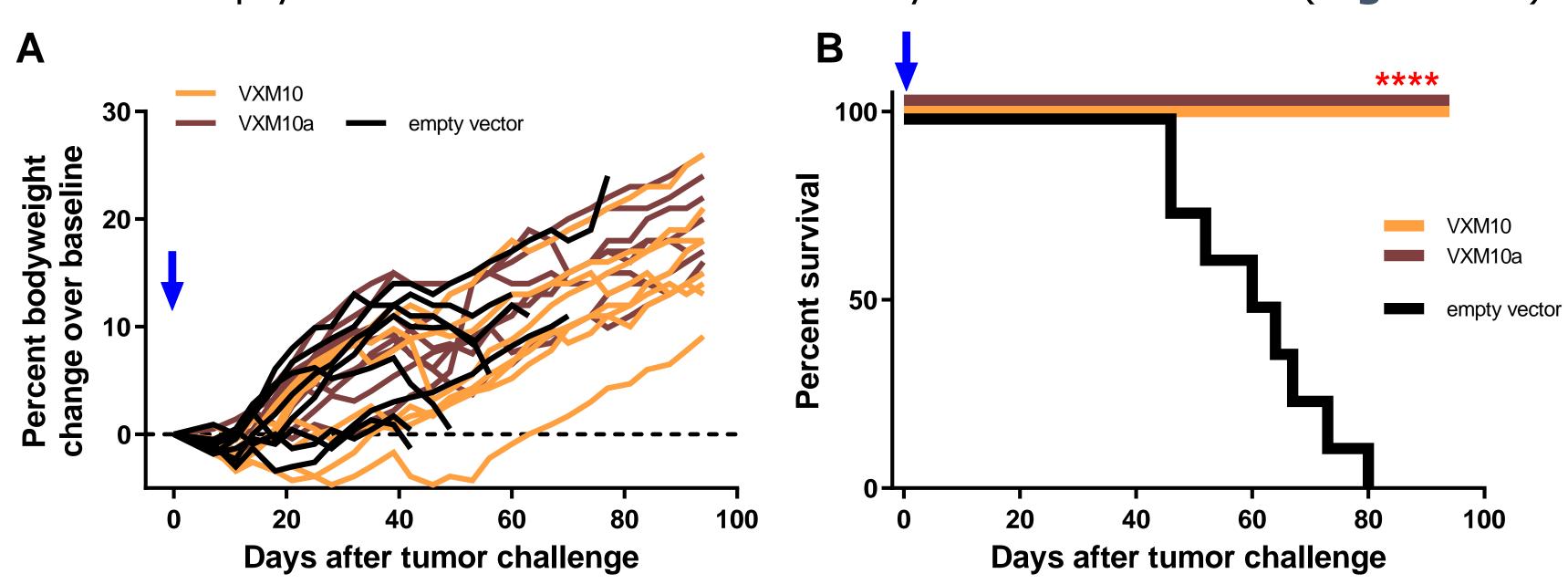


Figure 6. (A) Evolution of the bodyweight in each individual animal, and (B) overall survival in all treatment groups, in the therapeutic setting. The blue arrow represents the time point of FBL-3 challenge.

Antibody response

The systemic antibody response was evaluated by ELISA in the serum of animals vaccinated with either VXM10 or VXM10a, 79 days after the final vaccination (**Figure 7A**). Anti-PD-L1 antibodies were detected in a few animals vaccinated with VXM10 and VXM10a, and the response was more pronounced in the VXM10a/high-dose group, with 50% of the animals (3 out of 6) showing signal-to-background ratio above the cut-off value (**Figure 7B**).

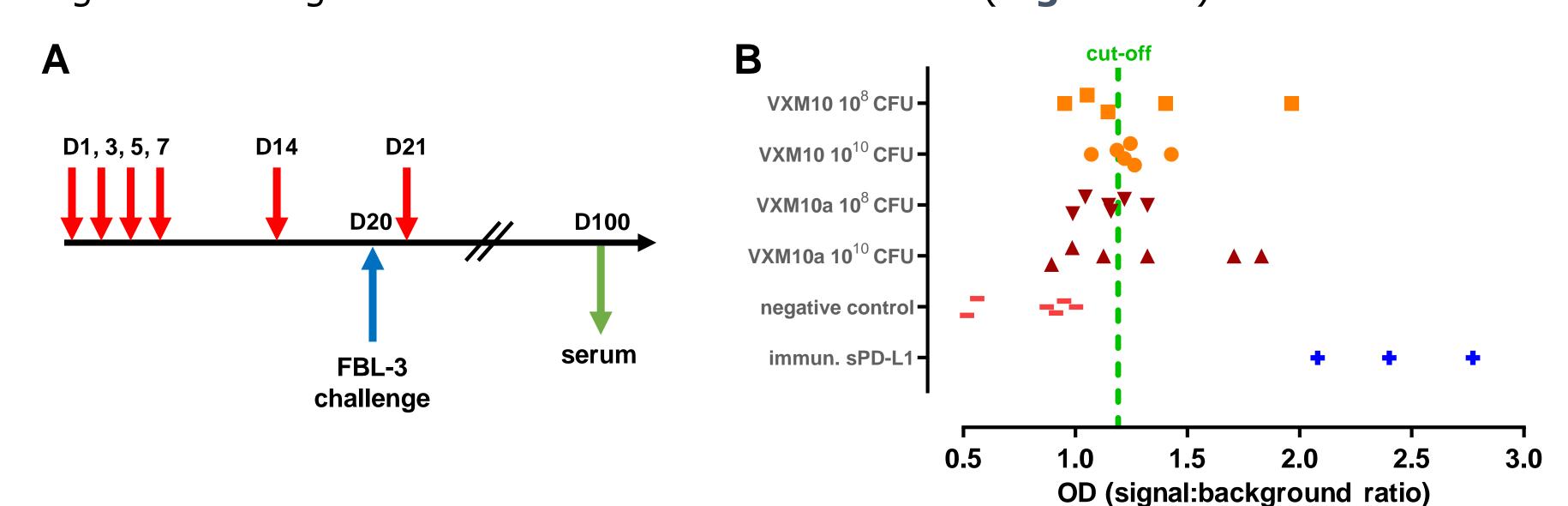


Figure 7. (A) Experimental design, and (B) anti-PD-L1 antibody response in sera collected 79 days after the final vaccination. The green dashed line represents the cut-off value (for 95% confidence). Soluble recombinant murine PD-L1 was used for immunization with CFA/IFA in the positive control group (blue).

Conclusions

- Prophylactic and therapeutic vaccinations with VXM10 and VXM10a induced a strong and sustained anti-cancer activity in the FBL-3 model of leukemia.
- This study provides evidence that VAXIMM's oral T-cell vaccination platform can be used to stimulate anti-tumor immunity against antigens of the immune checkpoint regulatory protein PD-L1.
- These data paved the way for advancing the clinical development of VXM10, in particular in leukemia.

References

1. Darji A. et al., Cell 1997; 91:765. 2. Niethammer AG. et al., Nature Medicine 2002; 8:1369. 3. Yamazaki T. et al., Journal of Immunology 2002; 169:5538.

Contact and Information

Heinz Lubenau, Ph.D. VAXIMM GmbH Chief Operating

Officer

MAFINEX-Technologiezentrum Office Julius-Hatry-Straße 1 Fax: 68163 Mannheim
☐ Marine Marine



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